II. CHEMICA

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NUTRITIONAL REQUIREMENTS OF LACTIC ACID BACTERIA

I. THE CALCIUM REQUIREMENTS OF STREPTOCOCCUS THERMOPHILUS STRAINS

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Nutritional requirements of lactic acid bacteria

From the standpoint of growth factor requirements lactic acid bacteria are among the most complex of the micro-organisms that have been investigated. The growth factors which either stimulate or are necessary for growth of lactic acid bacteria include various amino acids, peptides, vitamins of the B complex (and related compounds), purine and pyrimidine bases and their derivatives, and fatty acids. The known growth factor requirements of these bacteria have been discussed in numerous reviews ¹⁻⁷. Certain strains of lactic acid bacteria, including those which are commonly used in the dairy industry (e.g. the strains Streptococcus thermophilus, Lactobacillus helveticus, Lactobacillus lactis, and Lactobacillus bulgaricus) require unknown factors for growth in synthetic media. A more complete knowledge of the nutritional requirements of the lactic acid bacteria used in cheese manufacture is therefore a necessary foundation for any improvements in the quality of cheese.

Advantage can be taken of the fact that lactic acid bacteria require unknown organic growth factors to detect, isolate and identify such factors in biological materials. Until now, however, this possibility of discovering new growth factors by means of lactic acid bacteria used in cheesemaking has been very little utilized. The first aim of this work was to find basal media in which the growth of the lactic acid bacteria would be optimal and which could be used in attempts to isolate new growth factors. It was necessary, first of all, to find out in what way the different chemical compositions of the growth media affect the growth of lactic acid bacteria.

Experimental

Cultures and their maintenance

The experiments were carried out on 124 strains of lactic acid bacteria which had been isolated as pure cultures from various Finnish dairy products (milk, butter, buttermilk, cheese). The strains were kindly supplied by

Professor A. I. Virtanen, Director of the Biochemical Institute, Helsinki. These lactic acid bacteria represent a large group of bacteria whose nutritional requirements have not yet been investigated. Some of the strains are commonly used in the manufacture of cheese in Finland.

Cultures of the organisms were maintained as stab cultures in test tubes containing agar medium (TSHGA- agar medium) or sterile skim milk. The TSHGA- agar medium contained 1 per cent glucose, 0.5 per cent lactose, 0.5 per cent yeast extract, 0.5 per cent sucrose, 0.5 per cent Bacto-tryptone, gelatine 0.25 per cent, ascordic acid 0.07 per cent, and 1.5 per cent agar. The pH was adjusted to 6.7—6.8. The frequency of transfer varied from once a fortnight to once a month. The cultures were maintained at 4° between transfers.

Preparation of inoculum cultures

The composition of the inoculum medium was that described above but without the agar. This medium was divided into 7-ml aliquots which were sterilized in an autoclave 10-15 min. at 112° and then kept in a refrigerator. The inocula were prepared by transferring the organism from the stab culture to this medium. After being incubated at 37° for 12-48 hrs, depending on the strain in question, the cells were centrifuged and washed with 5-10 ml of 0.9 per cent sodium chloride solution. The centrifugation and washing were repeated three times. Finally the cells were suspended in 50 ml of saline. One drop of the slightly opaque suspension was used to inoculate the medium in each tube. Growth experiments were carried out at 37 or 42° . In general, the incubation was stopped when the logarithmic growth phase was reached in order to make possible a comparison of the rates of growth of different strains.

Compositions of basal media

The compositions of the basal media used in the investigation are shown in Table 1. All the media are modifications of the growth medium for lactic acid bacteria described by Anderson and Elliker 8. They were prepared by mixing stock solutions of the various compounds. Separate solutions of vitamins, purine and pyrimidine bases were prepared, but the inorganic salts were combined in a single solution unless stated otherwise. The basal media were adjusted to pH 6.7 before being used. All stock solutions were stored in a refrigerator and replaced monthly.

Component	Quant	ity	s	SII	SI	II SI	77	ві	-						
		<u> </u>		"		51	<u> </u>	ът	B		CI	C	$\Pi \mid C$	III	CIA
Glucose	mg	1	000	1000	100	0 100	1	000	1000			Ť	i i		
Lactose	»		500	500			- 1	500	1000	-\	000	100	1-0	00	1000
Sucrose	*	ľ	500	500			' '	500 500	500	1	00	50	- 1	00	500
DL-Aspartic acid	*		20	20	1	1	- (-		500	1. ~	00	50	0 50	00	500
L-Glutamic acid	*		10	10	1 -		1	20	20	- 1	20	20	0 2	20	20
Sodium acetate	l »	2				$\begin{vmatrix} 10 \\ 2000 \end{vmatrix}$		10	10	. 1	10	10) 1	0	10
Xanthine	*	-	2	2000	2000		1	-	1000	- 1	-	1000) 3	4	340
Casamino acids	*	110	- 1	1000		-	- 1	2	2		2	2	3	2	2
Tween 80	*		100	. 1		1	1	00	500	5	00	500	50	0	500
Oleic acid	, ,	'	1	100	100		1	00	100	10	00	100	10	0	100
DL-Tryptophan	, »		20	1	1		- 1	1	1	1	1	1	1	1	1
L-Cystine		-	- 1	20	20		2	20	20	2	0	20	4	- 1	20
Thymine	»	١.	10	10	10			10	10	1	0	10	- 1	- 1	10
Cytosine	μg	- 1	00	100	100		10	00	100	10	0	100	100		100
Hypoxanthine	*	j j	00	100	100	100	10	00	100	10	0	100	100		100
Adenine, Guanine, and	, »	1	00	100	100	100	10	00	100	10	0	100	100		100
Uracil, each	1		- 1		1		1					200	100	' ·	100
Ascorbic acid	mg	-	2	2	2	2		2	2		2	2	2		•
Thiamine HCl	*	- 1	10	40	20	20	2	0	20	2		20	20	- 1	2
iboflavin	μg	10	- 1	100	50	50	5	0	50	5		50	1	- 1	20
	*	10	00	100	50	50	5	0	50	50	1	50	50	- 1	50
Nicotinic acid	»	10	00	100	50	50	5	- 1	50	50	1	50	50	í	50
P-Aminobenzoic acid	$\mu \mathrm{g}$	10	00]	100	50	50	50	- 1	50	50	1	50 50	50	- 1	50
a-pantothenate	*	10	0]	100	50	50	50	- 1	50	50			50	- 1	50
Biotin	$m\mu g$	10	0 1	00	50	50	50	- 1	50	50	- 1	50	50	- 1	50
7itamin B ₁₂	»	2	2	22	11	11	11	1	11			50	50	I.	50
olic acid	»	100	0 10	00 8	500	500	500	i	500	11	1.	11	11	1	11
yridoxal phosphate	*	40		- 1	- 1	200	200	"	200	500	- 1	500	500	50	00
yridoxaminephosph.	$\mu { m g}$	1	1	10	5	5	2 00	1 -		200	2	200	200	20	00
yridoxamine HCl	»	10	. 1	10	5	5	5	ſ	5	5		5	5		5
yridoxal	*	20	1	- 1	10	10		- 1	5	5		5	5		5
yridoxine HCl	$\mathbf{m}\mathbf{g}$	4	1 '	4	2	2	10	1	10	10		10	10	1	0
$\mathrm{H_{2}PO_{4}}$	»	1000			-	. 4	2		2	2	1	2	2	1	2
${ m eSO_4}$. 7 ${ m H_2O}$	$\mu \mathbf{g}$			00 13	20	100	400	١.	-		1	-	12	-	-
nSO_4 . $4 H_2O$	mg	13		3	. 1		400	ı	00	400	- 1	00	400	40	0
gSO_4 . $7 H_2O$	»	80	1 -	· . I	3 40	0.4	0.	- 1	0.4	0.	1	0.4	0.4		0.4
Cl	»		١	1		30	30	J	30	30	:	30	30	3	0
aCl	" »	4	_	- 1	50 1	40	140	14	10	140	14	10	140	14	0
ycero-2-phosphate	»	4		4 -		-		-	-	4	1	4	4	1	4
dium citrate . $2 H_2O$	»	E00	_	1	78	-	156	15	66	_	-	-	468	468	- 1
ccinic acid	*	500	-	-	.	-		-	-		-	_			-
	"			1 -	.	- 1		1 -	_		1	- 1	295	295	-

Medium SIIIb is the same as SIV but contains also 156 mg of glycero-2-phosphate per $100\,$ ml.

All media contained traces of ammonia salt because xanthine was dissolved in strong ammonia.

Sodium hydroxide or hydrochloric acid was used to neutralize the media to pH 6.8 and therefore the concentrations of $\rm Na^+$ or $\rm Cl^-$ ions in the media are somewhat higher than the table indicates.

General procedure

In principle the experimental procedure used in the study was the same as that used in this laboratory for the determination of vitamins and amino acids. Two and a half millilitres of the basal medium was added to a test tube and diluted with water or supplements to 5 ml. The test tubes were then plugged with cotton wool and autoclaved 5—10 min. at $112-115^{\circ}$. After being cooled, the tubes were inoculated and then incubated at 37 or 42°. The growth of the lactic acid bacteria was followed turbidimetrically, the extent of growth being recorded as galvanometer readings of a Klett-Summerson colorimeter with a 590-660 m μ filter. In the tables showing the results, the rates of growth are expressed directly as galvanometer readings.

Results

In the initial stage of the work, several of the 124 strains of lactic acid bacteria tested appeared to possess very complex growth factor requirements. It also became clear during the first experiments that some strains of Streptococcus thermophilus require calcium for growth when synthetic media are used. Because it is very important to know the exact nutritional requirements of these calcium-requiring strains, the next step was a detailed investigation of the effect of different inorganic and organic components of the basal media on the growth of these bacteria. The results of these experiments are shown in Tables 2—25. It should be noted that these results relate to strains which are sensitive to calcium. About 30 strains (mainly Streptococcus thermophilus) of the 124 strains preliminarily tested were sensitive in this respect.

Potassium. The effect of potassium on certain strains is shown in Table 2.

Table 2.
Effect of potassium concentration.

Basal medium	Strain		Incubation					
		0	0.7	1.4	2.1	2.9	4.3	at 42°C, hours
SIII	Str 10		150	110	1			
»	Str 11			110	40	_		24
		_	110	100	90	60		24
»	Str 110	_	150	110	90	_		24
**	Str 111		170	160	160	140	90	
»	Cas I	_	150	170			20	24
»	Cas J (7)				130	60	-	70
»		-	200	230	325	_		70
	Kungsbölen k.		190	190	190	180	150	24
»	Kauhajoen k.		190	170	120		-30	24 24

As can be seen, potassium ions are required by all the strains. Already the lowest concentration of potassium (37 μ moles per ml) effected maximal growth under the experimental conditions.

Manganese. The effect of manganese on the growth of 10 strains is seen in Table 3. The basal media used in these experiments were SII and SIII, but without the manganese sulphate. The results show that manganese ions are not generally required by these strains.

Table 3.
Effect of manganese concentration.

Basal medium	Strain			μ g	Mn+	-/ml			Incubation	
		0.0	1.0	3.0	5.0	7.5	12.5	25.0	at 42°C, hours	
» »	Str 10 Str 10 Str 10 Str 11 Str 110 Str 111 Kauhaj. k. Kungsb. k. Cas J (7) Cas I	125 180 75 55 155 175 179 210 20 146 145 175 235 —	77 77 160 172 174 201 144 170	82 76 148 168 172 198 115 146	130 180 88 79 146 162 160 195 70 165 125 215 290 115 115 183	91 79 142 161 178 200 57 65 75 280 125 126 185	100 170 90 79 140 168 165 191 29 54 25 260 98 120 166	50 33 105 108 110 179	18 40 24 24 24 24 24 24 24 24 24 40 18 24 40 60	

Iron. Table 4 shows the effect of iron on the growth of 9 strains. Iron (II) ions do not seem necessary for rapid growth under the experimental conditions used. With some strains, iron was observed to promote growth.

Magnesium. The magnesium requirements of 10 strains were tested. As can be seen from Table 5, magnesium ions are essential for the growth of most strains and stimulate the growth of the other strains.

Citrate. The preliminary experiments showed that sodium citrate inhibits the growth of certain strains of lactic acid bacteria (named Group II). These findings were confirmed and the experiments were extended to an

Table 4. Effect of iron concentration.

Basal	Strain		:	με	Fe++	/ml			Incubation at 42°C,
medium	Suranı	0.0	0.8	1.6	3.2	4.0	4.8	8.0	hours
SIII	Str 10	57	67	54	40		35	26	24
SII	Str 10	110	158	188	155	142		90	24
»	Str 10	140	165	198	150	160	ĺ	180	45
SIII	Str 11	38	69	63	58		50	74	24
SII	Str 111	210	165	250	190	186		180	45
\mathbf{SIII}	Str 111	155	148	157	145		136	136	24
*	Kauh. k.	165	169	161	161		144	122	24
*	Kungsb. k.	178	181	172	172		150	150	24
*	Str 110	138	132	119	113		_	_	24
*	Cas J (7)	10	96	115	100		117	95	24
»	Cas I	145	187	187	199	·	170	132	24
SII	» I		97	135	105		110	114	27
· »	» »		115	115	97		78	50	45
* *	Lb helv.					-			
	1182	20	78	129	206		200	190	45

Table 5. Effect of magnesium concentration.

Basal	Strain			μg Mg	;++/ml			Incubation at 42°C,
medium	 	0.0	20.0	40.0	60.0	80.0	120.0	hours
		-						
SII	Str 10		125	125	120	107		27
SIII	Str 10	_	105	106	103	90	95	24
SII	Str 10		153	174	178	178		40
SIII	Str 11	47	106	112	100	100	92	24
»	Str 111		170	167	167	165	160	24
»	Kauh. k.	-	170	166	155	152	145	24
»	Kungsb. k.	_	194	195	198	206	209	24
»	Cas J (7)		115	230	226	210	140	72
»	Str 110		128	111	138	46	11	24
SII	Cas I	_	15		10	55		24
»	» »	30	230	220	270	185		40
SIII	» »	12	185	192	189	174	173	72
SII	Ylilied. k.		130	135	120	110		24
»	» »	167	184	185	166	179		40
»	Lb helv. 1182	5	24	16	14	47		64

examination of the tolerance of citrate by 25 strains of Group II using basal medium SIII (or SII). The citrate tolerance varied considerably with different strains. The strains were divided into four groups according to the degree of tolerance (Table 6).

Table 6.

Tolerance of citrate by different strains of lactic acid bacteria.

Group IIA,	Group IIB, tolerates $1-2~{ m mg}$	Group IIC,	Group IID,
tolerates less		tolerates	tolerates more
than 1 mg		2-3 mg	than 3 mg
Str 11 Str 40 Str 75 Str 100 Str 102 Str 103 Str 110 Ths¹) Kauhaj.k. Ylilied.k.²)	Str 10 Str 11R Str 72 Str 74 Str 101 Str 1111 Cas I ²) Cas J (7) H.th.str	Hatt.s. Lb lact 1183 B.c.h.H.	Lb 80 Lb 83 Lb helv 11821)

¹⁾ Strain tested in medium SII.

Table 7.
Effect of calcium.

Strain	<u> </u>				μg Ca	++/ml				
		1	2	3	4	5	10	15	20	30
Str 10 Str 10 Str 11 Str 11 Str 11R Str 11R Str 110 Str 110 Cas I " "	5 3 1 6 9 7 - - 39 40	18 14 10 19 18 19 15 14 —	6 -12 5 45 110 18 92 39 65	2 6 11 3 82 100 178 175 63 66	95 -6 6 105 135 142 184 94 97	128 115 11 7 108 140 175 182 117 127	125 135 22 11 142 139 230 225	52 41 140 137 200 214 195 200	130 125 93 26 141 139 228 200 240 240	130 140 114 85 139 140 225 190 244 252

Basal medium S, in which the concentration of sodium citrate was 2.5 mg per ml, was used. Incubation 48 hrs at 42° .

²⁾ Strain tested in media SII and SIII.

Calcium. In a medium containing citrate, calcium is either necessary for or a stimulus to the growth of most strains. In a citrate-free basal medium, calcium has no effect on the growth in most cases (Table 8). However, as

Table 8.

Effect of calcium on growth in media containing citrate (SIVSS) and in citrate-free media (SIV). Incubation at 42°.

					1	1				
			SIVSS					SIV		
Strain	ŀ	$\mu g C$	a++/m	1	Hours		μg C	a++/m	1	Hours
	0	5	10	15	-		T _	T	Т	- Hours
	<u> </u>	1 "	10	13	1	0	5	10	15	
Str 10			30	90						
Str 10R		-	30	30	e 24	50	50	45	45	o 24
Str 11		35	45	4-	72		15	40	50	e 24
Str 11R		35	45	45	e 24	50	50	55	55	o 24
Str 12	-	-	40	_	72		50	40	40	e 48
Str 40	-	40	40	80	e 72		15	45	60	e 24
Str 72		40	50	60	e 48	60	60	60	65	o 48
Str 74	-	-	90	100	e 24		80	100	105	e 24
Str 75	_	50	80	75	e 48	70	70	70	75	o 24
Str 75	_	40	45	55	e 48	70	60	. 60	70	o 48
	_	_	75	130	e 72		30	60	80	e 24
Str 100	_	-	45	45	e 48	55	60	65	60	o 24
Str 101	-	-	50	30	e 48	_	5	20	20	e 24
Str 102	_	20	60	60	e 48	35	45	45	50	o 24
Str 103	-	<u> </u>	60	65	e 48	40	55	60	55	o 24
Str 110	<u> </u>	80	145	115	e 24	155	120	-130	135	o 24
Str 111		80	120	105	24	140	125	120	105	o 24
\mathbf{Th}		-	45	80	e 24	70	105	110	115	s 24
Ths	_	5	30	105	e 24		50	110	110	e 24
Cas I		40	100	110	e 24	110	125	125	120	0 24
Cas J (7)	5	30	100	140	s 24	130	140	140	140	0 24
H.th.str.	_		10	5	e 48	_	10	30	30	e 24
Hatt.s.	20	55	75	75	s 24	140	125	120	105	o 24
Kauh.k.		130	150	160	e 48	145	160	155	155	0 48
Kungsb.k.	110	50	100	130	48	120	120	130	120	0 48
Lb 83	135	135	125	100	o 48	150	130	125	130	
Lb lact. 1183	10	130	125	105	s 48	125	130	135	135	0 48
			120	100	9 40	120	190	135	139	s 48

Notes: e = calcium essential for growth.

s = calcium stimulates growth.

o = calcium has no effect on the growth.

can be seen from Tables 9 and 12, the composition of the basal media also otherwise affects the calcium requirements of the strains. On the basis of their calcium requirements in the SIV and SIVSS basal media, the strains were divided into three groups as shown in Table 10. These groups are:

Table 9. Effect of calcium on the growth in media SII and SIIIb lacking citrate. Incubation at 42°.

Strain	με	SIII g Ca+	b +/ml		Hours		μ_i	S g Ca	II - - /m	l		Hours
	0	5	10	15		0	5	10	15	20	25	
G. 10	50	70	65	65	0 24	125	115	110	105	105	105	0 24
Str 10	30	5	30	45	e 24	120	110					
Str 10R Str 11	45	60	60	60	0 24	80	80	80	80	90	80	o 24
	45	55	55	60	0 24	110	105	100	90	90	90	o 24
Str 11R Str 12	40	5	20	35	e 24							
Str 12 Str 40	70	70	70	70	0 24	100	115	110	110	110	110	0 24
Str 40	25	120	130	130	s 24	_	20	30		100	100	e 24
Str 74	85	85	75	75	0 24	_	120	120	120	120	125	e 24
Str 75	60	65	60	55	o 24	_	20	100	80	70	.110	e 24
Str 77	_	20	65	85	e 24			_		5	15	e 24
Str 100	70	70	70	70	o 24	_		5	20	25	20	e 24
Str 101	40	50	50		o 24		20	35	55	50	50	e 24
Str 102	25	50	55	60	s 24	_	_	5	5	5	10	e 24
Str 103	45	70	70	65	o 24	_	5	10	35	50	45	e 24
Str 110	160	120	120	125	o 24	140	155	155	165	160	160	o 24
Str 111	145	130	135	135	o 24	190	205	205	210	210	215	o 24
Th	5	10	115	140	s 24	_	15	100	135	150		e 24
Ths	150	160	175	190	s 24	-	35	125	145	150	155	
Cas I	20	125	125	125	s 24	185	290		320			s 24
Cas J (7)	170	180	200	195	o 24	-	140	180	250	290	270	1
H.th.str	_	45	60	60	e 24	_	-	5	5	5	10	e 24
Hatt.s.	180	180	190	180	o 24							
Kauh.k.	125	125	135	130	0 24	185	190	190	190	190	185	ł
Kungsb.k.	135	140	140	145	o 24	185	185	185	185	185	175	1
Ylilied.k.	45	45	35	35	o 24	-	-	40	50	60	65	1
Lb 80	175	175	190	170	0 24	220	220	250	245	1	225	1
Lb 83	105	105	100	105	o 24	320	310	330	320		320	
Lb helv. 1182						200	205	150				0 2
Lb lact. 1183	90	165	195	200	s 24	50	80		130	140		s 2

Notes: e = calcium essential for growth.

s = calcium stimulates growth.

o = calcium has no effect on the growth.

Ha: strains requiring calcium ions in both media,

IIb: strains requiring calcium ions only in the medium containing citrate, and

IIc: strains not requiring calcium ions in either medium.

In addition, the first two groups can be divided into subgroups.

Hal and Hbl: calcium ions are essential for growth, and

IIa2 and IIb2: calcium ions stimulate the growth.

Similar groups can be formed on the basis of the experiments carried out in the basal media SII and SIIIb — these media differ from each other primarily in phosphate content (Table 11). The groups are in this case:

IIx1: the strains require calcium ions in both media,

IIx2: calcium ions stimulate the growth in SIIIb medium,

IIy2: calcium ions are not necessary for growth in medium SIIIb but stimulate the growth in medium SII, and

IIz: the strains do not require calcium ions for growth in either medium.

These results show that the importance of calcium ions is not restricted to a counteraction of the growth inhibition by citrate. It seems probable that phosphate in the concentrations employed also exerts an inhibiting effect on several strains. Calcium ions seem to counteract this inhibition as well.

The experiments proved that calcium counteracts the growth inhibition by citrate. The next step was to study possible counteracting effects of the other components of the basal media.

Table 10.

Grouping of strains according to the tests carried out with calcium in media SIV and SIVSS.

IIa1	IIa2	IIb1	IIb2	IIe
Str 10R Str 11R Str 12 Str 72 Str 77 Str 101 Ths H.th.str.	Th Lb lact 1183	Str 10 Str 11 Str 40 Str 74 Str 75 Str 100 Str 102 Str 103 Str 110 Str 111 Cas I Kauhaj.k.	Cas J (7) Hatt.s.	Kungsb.k. Lb 83

Table 11. Grouping of strains according to tests carried out with calcium in the media SII and SIIIb.

IIx1	IIx2	IIyl	IIy2	IIz
Str 10R Str 12 Str 77 H.th.str.	Str 72 Str 102 Th Ths Cas I Lb lact 1183	Str 74 Str 75 Str 100 Str 101 Str 103 Cas J (7) Ylilied.k.		Str 10 Str 11 Str 11R Str 40 Str 110 Str 111 Hatt.s. Kauh.k. Kungsb.k. Lb 83

Table 12. Effect of calcium in the medium CIV containing 2.5 mg of sodium citrate per ml. Incubation 45 hours at 42°.

Strain	μ g Ca ⁺⁺ /ml:		25 6	50 12	75 19	100 25
Ths			50	88	98	90
Str 12		_	-		_	
Str 10		_	_	5	62	71
Th	5.00	_	_	13	13	87

Phosphate. In the experiments concerning calcium requirements it appeared that high phosphate concentrations inhibited the growth of several strains. Phosphate seemed to have no effect on the growth in calcium-free media when used in low concentrations, but inhibition of growth was soon observed when the concentration was increased. Calcium ions counteracted this inhibition (Tables 13, 14 and 15).

Acetate. As can be seen from Table 16, a much stronger growth was observed in a medium containing both calcium and acetate ions than in a medium containing acetate ions, but no calcium.

Table 13. Effect of phosphate.

CICaTa	ı	n -	μmole/ml										Incub.
CICaTa			0	15	20	30) 4	40	60	75	8	0	hours
100010	1) Str 1(_	_			İ	Ť		 	+-	+-	
CIITa ²)	Str 10)	80	_	70	_					-	- 2	4 K4
CII	Str 10)	40	- 1	30		1	50	40	10		24	
CIIB3)	Str 10	- 1	80	- 1	80		- 1	10	-	_		24	
CI	Str 10		_		48			70	60	40		24	
»	Str 10	- 1	5		141		1	28	14		1	9 48	1
CIV	Str 10		_		141		20	- 1	-1		-	- 48	
»	Str 10	- -:	30		_		1	6	6		(6 48	
CI	Str 74		_		4		1 -	-	-		-	- 48	, .
»	Str 74	-	_		59		1	3	11		15	5 48	»
\mathbf{CIV}	Str 74	17	9	- 1	38		-	-	-		-	48	»
»	Str 74	1 -	_	.	_		-	i	-			48	*
CI	Str 75	-	0	1	3		-		-		_	48	»
*	Str 75	- 1	5	,	09		4:	3	13		13	48	»
CIV	Str 75	- -	_	1	09			- -	-	- 1		48	
CII	Str 77	7	n l		70		-	i	-	- 1	7	48	l »
CIIB³)	Str 77		_	i	10	- 1	40) 1	0	-		24	K4)
ZI .	Str 101	1 _	.	Ι,	37			-	-		_	24) »
»	Str 101	9)	10		- 1		22	$2 \mid$	- 1	22	48	Na5)
IV	Str 101	_		1	5	1	_	-	-	. 1	-	48	»
ICaTa¹)	Hatt.s.	10	1 _	-	9			1 -	-	- 1		48	»
$IITa^2)$	Hatt.s.	170	1	23	0	-	200	-	- .			24	K4)
II	Hatt.s.	170		27			200	160		50		24	*
$IIB_3)$	Hatt.s.	190		29			280	270	1	50	1	24	»
Į.	Cas J (7)	68		13	- 1	-	280	280	, -	80		24	»
	Cas J (7)	80		19		- 1	222	278	1		22	48	Na5)
\mathbf{V}	Cas J (7)	270		300	- 1			-		.		48	*
1.	Cas J (7)	261		000			300	7	1		6	48	»
CaTa¹)	Lb 80	120	210		19	0	_		1		- j	48	»
ITa²)	Lb 80	150		300		- 1	200	90	1	-		24	K4)
I	Lb 80	150		300		. 1	300	300	30			24	»
IB³)	Lb 80	150	.	300	- 1	- 1	300 300	300 300	30			24	»

¹⁾ CI + 5 μg Ca++/ml + 100 μ mole tartaric acid/ml.

²⁾ CII + 50 μ mole tartaric acid/ml.

s) CII + 50 μ mole/ml each of malic acid, malonic acid, succinic acid, and tartaric

⁴⁾ The phosphate was added in the form of primary potassium phosphate.

⁵⁾ The phosphate was added in the form of primary sodium phosphate.

Table 14. Effect of potassium phosphate in medium CI containing acetate, glycerophosphate,

succinate and tartrate (5 μ mole/ml of each) tested with and without 5 μ g Ca⁺⁺/ml. Incubation at 42°.

Strain			$\mu\mathrm{mol}$	e/ml			Incubation
	0	5	10	20	50	100	hours
Without calcium:							
\mathbf{Th}	72	78	62	48	12		24
Str 77	70	57	61	85			
Str 10	50	55	38	35	11	7	70
Hatt.s.	107	94	88	79	79	61	24
Lb. 80	-	184	194	181	194	190	24
With calcium:				101	101	190	70
\mathbf{Th}	134	129	145	120	100	49	94
Str 77	64	76	82	79	62	22	24
Str 10	66	58	63	49	34	5	24
Hatt.s.	92	103	137	114	140	$\frac{3}{73}$	24
Lb 80	175	181	195	199	220	198	24 70

Malate, malonate, succinate, tartrate and glycerophosphate. As very little information is available on the effect of these organic acids on the growth of lactic acid bacteria, a series of experiments were carried out with these acids. The preliminary findings are shown in Tables 17—23. These findings still require checking, but they show clearly that succinate in low concentration stimulates the growth of certain strains in the absence of calcium ions but stimulates the growth even more when calcium ions are present.

Table 24 shows results obtained in experiments where the effect of citrate was studied in a basal medium lacking acetate but containing other organic anions. Also these findings are preliminary, but it is seen that growth inhibition by citrate is stronger when organic anions other that acetate are present in the medium.

Other organic compounds. Table 25 shows results obtained in experiments with the following compounds: oxalacetate, α -ketoglutarate, fumarate, formate, glycerol, lactate, pyruvate and propionate. It is not possible to draw any definite conclusions from these results, but it is interesting to note that oxalacetate inhibited the growth of Streptococcus thermophilus strains, and that calcium ions counteracted this inhibition to a certain extent. This particular finding has been confirmed subsequently.

The promotion of growth by calcium and other chemical compounds in the media is being investigated. The experiments carried out to date have shown that no reciprocal influence exists between calcium and certain sugars such as sucrose, glucose and lactose. It was found that some strains of *Streptococcus thermophilus* cannot utilize glucose; all the strains utilized sucrose and lactose.

Table 15.

Effect of calcium in media containing sodium phosphate. Incubation at 42°.

Basal medium	Phosphate	Strain		μ	g Ca+-	⊦/ml		Incubation
medium	μmole/ml		0	5	10	20	40	hours
CI	50	Str 10	78	173	141	1		1
»	50	Str 10	206	131	141 128	149	142	42
»	75	Str 10	260	288	241	130	153	42
CIV	50	Str 10	200	200	6	162	216	42
»	75	Str 10	_		0	10	33	48
CI	50	Str 74	10	10	86	5	5	48
*	50	Str 74	10	208	273	177 246	256	48
»	75	Str 74	315	220	220	250	238	48
\mathbf{CIV}	50	Str 74	010	220	220	1	280	48
»	50	Str 74				10	31	48
»	75	Str 74					11	48
CI	50	Str 75	12	185	32	26	200	48
»	50	Str 75	131	226	206	214	262	48
»	75	Str 75	101	62	220	1	210	48
CIV	50	Str 75	5	02	5	208	167	48
»	50	Str 75	,		Ð	17	33	48
»	75	Str 75					11	48
CI	50	Str 101	60	134	190	100		48
»	50	Str 101	85	100	136	133	134	48
»	75	Str 101	09	18	105	123	123	48
CIV	50	Str 101	_	18	216	6	124	48
»	50	Str 101	_	_	6	38	35	48
» »	75	Str 101	1		_	-	9	48
CI	50	Cas J (7)	236	940	100	-	_	48
»	50	Cas J (7)	212	240	109	140	141	48
»	75	Cas J (7)		220	202	153	111	48
CIV	50	Cas J (7)	258	296	298	280	125	48
»	50	Cas J (7)	5	281	299	330	335	48
»	75			-	8	16	20	48
"	10	Cas J (7)		-				48

Table 16.

Effect of sodium acetate in medium CI containing glycerophosphate, potassium phosphate, succinate, and tartrate (5 μ mole/ml of each), tested with and without calcium. Incubation at 42°.

$\mu \mathrm{g}$ $\mathrm{Ca^{++}/ml}$	Strain			μ mol	m le/ml		,	Incubation
/1111		0	5	10	20	50	100	hours
5 - 5 - 5 - 5	Th Th Str 77 Str 77 Str 10 Str 10 Hatt.s. Hatt.s. Lb 80 Lb 80	31 124 - 5 43 60 51 67 142 172	79 144 78 75 50 68 92 105 177 180	94 149 76 76 53 65 85 77 222 208	97 163 92 92 57 73 58 70 228 246	101 167 87 87 70 78 68 54 258 272	149 7 7 66 82 85 61 294 284	24 24 70 24 24 24 24 24 70

Table 17.

Effect of malic acid. Incubation 48 hours at 42°.

Basal medium	Strain		μ mo	ole/ml	
medium		0	50	100	150
CICa ¹)	Th	20			
${f BI}$	Th	30	10	-	
\mathbf{BII}	Th	90	_	_	-
CICa	Str 77	10	10	_	
BI	Str 77	20	10	_	_
BII	Str 77	_	_	_	
$CICa^1)$	Hatt.s.	80	170	170	20
BI	Hatt.s.	70	10	_	_
BII	Hatt.s.	240	200		
CICa	Lb 80	70	120	70	100
BI	Lb 80	70	220	230	200
BII	Lb 80	400	230	180	190

Table 18.

Effect of malonic acid. Incubation 48 hours at 42°.

Basal	Strain		μ	mole/m	ı	
medium	Solum	0	50	100	150	200
BI	\mathbf{Th}	30			_	
BII	\mathbf{Th}	10	'		_	_ ·
BI	Str 77	20	· ·		_	
BII	Str 77	40	_		_	
BI	Str 10	50	20		_	
BII	Str 10	70	_		_	
BI	Hatt.s.	170	30		_	
BII	Hatt.s.	240	200			
BI	Lb 80	80	220	180	50	10
BII	Lb 80	260	240	190		

Table 19. $Effect \ of \ high \ succinic \ acid \ concentrations. \ Incubation \ 48 \ hours \ at \ 42^{\circ}.$

Basal	Strain			μ mole/	ml	
medium	Strain	0	50	100	150	200
CICa1)	\mathbf{Th}	10	90			
BI	${f Th}$	40	_	_	_	l
BII	${f Th}$	_	_	_		l
CICa	Str 77	10	150	_	_	
\mathbf{BI}	Str 77	20		_	_	
BII	Str 77	_	_			
CICa	Str 10	10	120	50		_
BI	Str 10	20	140	200		10
BII	Str 10	60	_	_		_
CICa	Hatt.s.	80	170	170	20	
BI	Hatt.s.	100	210	10		
BII	Hatt.s.	220	180	-		-
CICa	Lb 80	70	90	120	110	20
BI	Lb 80	80	200	180	160	30
BII	Lb 80	260	210	150	140	

Table 20.

Effect of low succinic acid concentrations in medium CI containing acetate, glycerophosphate, and potassium phosphate (5 μ mole/ml of each), tested with and without Ca++. Incubation at 42°.

$_{ m Ca^{++/ml}}^{ m \mu g}$	Strain		Incubation					
,		0	5	10	20	50	100	hours
10 	Th Th Str 77 Str 77 Str 10 Str 10 Hatt.s. Hatt.s. Lb 80 Lb 80	78 115 26 67 51 54 89 95 133 135	94 139 14 79 55 65 99 88 191 187	95 157 — 82 72 71 125 108 200 224	95 169 — 87 82 98 76 65 226 248	196 	20	24 24 24 24 24 24 24 24 24 70

Table 21.

Effect of tartaric acid. Incubation at 42°.

Basal medium	Strain				μ r	nole/n	nl			Incubation
		0	5	10	20	5 50	0 10	00 15	50 200	house
BI	Th	4					İ		1	
BII	Th			- 1	- 1	-	-	·	.	48
$CICa^{1}$)	\mathbf{Th}	20				-		-	.	48
CIb	\mathbf{Th}	96	1			. -	-	-	1 -	48
${f CIbCa}$	Th	118	1	1 0	'	1	-	1		24
\mathbf{BI}	Str 77	10	~0*	2 14	1 142	2 7	9 -	1		24
BII	Str 77				1	-	-	1 -	. -	48
CICa	Str 77	10	1			-	-	-	-	48
CIb	Str 77	59	1			110) -	-	-	48
CIbCa	Str 77	98	101	1		-	-	_	1	24
BI	Str 10	30	101	100	78	1	-			24
BII	Str 10	60				10	-	-		48
CICa	Str 10	10				l -	-	-	1 -	48
CIb	Str 10	66	63	0.5	1	30	-	-	-	48
CIbCa	Str 10	63	77	65	54	_	-	-		. 24
BI	Hatt.s.	100	"	63	65	_	-		1 1	24
BII	Hatt.s.	210				210	10	-	-	48
CICa	Hatt.s.	10			l	190	_		-	48
CIb	Hatt.s.	130	140			40	-	<u> </u>	_	48
CIbCa	Hatt.s.	131	142 126	142	130	71		_		24
BI	Lb 80	80	120	117	123	93	25			24
BII	Lb 80	260				200	180	160	30	48
CICa	Lb 80	70			1	200	170	130	10	48
CIb	Lb 80	158	176	100	200	120	120	130	10	48
CIbCa	Lb 80	159	183	185	202	216		1		24
		100	100	206	218	234	185	- 1	- 1	24

Table 22. Effect of glycero-2-phosphate in medium CI containing 5 μ mole/ml each of acetate, potassium phosphate, succinate, and tartrate. Incubation at 42°.

$\mu_{ m g}$	Strain			μ m	ole/ml			Incubation
		0	5	10	20	50	100	hours
10 10 10 10 10	Th Th Str 77 Str 77 Str 10 Str 10 Hatt.s. Hatt.s. Lb 80 Lb 80	34 94 26 68 23 44 98 87 —	87 139 11 86 50 69 101 98 178 198	97 156 — 100 82 91 74 53 214	194 	18 — 18 — 71 210 34 28 280		24 24 24 24 24 24 24 24 24 70

Table 23.

Effect of malic acid, malonic acid, succinic acid, and tartaric acid in medium CI.

Incubation 48 hours at 42°.

Strain	mali	c acid, ma tartaric ac	alonic acidid, μ mole	l, succinic /ml of ea	acid, ch
	0	12.5	25	37	50
Th		20	50	50	60
Str 77	_	10	40	50	
Hatt.s.	50	90	90	130	120
Lb 80	60	130	160	160	100

Table 24.

Effect of sodium citrate in the media CI, CII, and CIV. Incubation 48 hours at 37°.

Basal medium	Strain					$\mu\mathrm{m}$	ole/ml	Į			
		0	2	4	10	15	20	27	34	48	6
CI	\mathbf{Th}	61	20	34					İ	1	
CII	\mathbf{Th}		1	1	-	-	-	-	-	-	_
CIV	Th	109	95	48	-	-		_	_	_	_
CI		186	176	-	_	-		-	_	_	l _
CII	Str 77	-	-	-	-	-		_		_	
1	Str 77	126	95	80	_	_		_			
CIV	Str 77	138	89	-	_	_	_	_		_	
CI	Str 10	-	17	21	_	l			_	_	_
CII	Str 10	67	59	68		_		-	_	_	
CIV	Str 10	164	142	87			_	_		_	
CI	Hatt.s.	76	98	126	128	49	100	-	-		
CII	Hatt.s.	193	192	183	6	49	120	-		-	_
CIV	Hatt.s.	272	180		-		_			-	
CI	Lb 80	65		127	7		_		-	_	
CII	Lb 80	1	88	133	141	164	138	129	26	-	
CIV		176	142	139	122	43	4		- 1	_	
011	Lb 80	280	248	198	192	137	'	_	_	_	

Table 25.

Effect of oxalacetate (= Oa), α -ketoglutarate (= Kg), fumarate (= Fu), formate (= Fo), glycerol (= G), lactate (= La), pyruvate (= Py), and propionate (= Pr) in medium CI, tested with 10 μ g Ca⁺⁺/ml and without calcium. Incubation 12 hours at 37°.

Compound	Strain		w		t Ca+	-+.				-241- 4	O- 1-1-			
00			without Ca++, µmole/ml						with Ca++,					
On		0	5	10	20	50	100	0	5	10	20	50	100	
	Lb 80	103						1					†	
Kg	Lb 80	79	117	130	107	ŀ		108	3	-	-			
Fo	Lb 80	81	77	73	167 49	90		105	130	167	174			
Fu	Lb 80	73	150	150	124	30		106	100	79	1	59	46	
La	Lb 80	74	94	105	117	112		117	163	168	135	126	122	
Py	Lb 80	90	73	58	47	115	70	85	85	126	134	113	88	
Pr	Lb 80	83	125	143	146	43 212	20	109	89	78	99	93	38	
G	Lb 80	72	75	74	78	78	135	98	136	150	146	151	135	
Oa	Hatt.s.	80		14	10	10	82	97	95	94	96	87	89	
Kg	Hatt.s.	68	99	130	140			65	-		_			
Fo	Hatt.s.	74	68	66	65	60	45	73	100	137	127			
Fu	Hatt.s.	58	118	145	173	158	35	77	64	67	68	68	53	
La	Hatt.s.	66	78	86	86	21	35	80	131	155	173	161	146	
$\mathbf{P}\mathbf{y}$	Hatt.s.	71	69	69	58	46	36	75 69	82	87	90	27	8	
\mathbf{Pr}	Hatt.s.	67	97	123	153	98	20	71	72	63	62	52	37	
G	Hatt.s.	68	65	67	68	68	68	ŀ	103	123	134	149	25	
Oa	Str 10	23	49	_	00	00	08	$\begin{vmatrix} 68 \\ 31 \end{vmatrix}$	69	72	69	69	67	
Kg	Str 10	32	33	41	68			36	65	97	_			
- 1	Str 10	28	22	17	19	15	9	36	34	55	76			
. 1	Str 10	14	19	21	36	24	9	35	30	19	20	12	9	
1	Str 10	29	22	16	13	24		34	38 32	44	57	73	64	
1	Str 10	17	18	24	40	86	40	28	39	24	20	_		
	Str 10	14	36	47	47	19	4	26	46	53	67	84	45	
G	Str 10	13	13	15	16	13	18	24	20	49	56	19	8	
Oa	Th	16	_	_	_	10	10	33	75	18 59	21	21	22	
	Th	26	40	75	68			37	54	74	-	.		
	Th	21	31	29	25	14	7	37	47	40	91	10		
1	Th	10	13	15	16	28		31	4/	40	28	18	14	
	Th	24	19	22	15			43	49	29	_	5		
	Th	15	15	78	33	97	117	T0	40	4	4 3	_	-	
- 1	Th	14	29	31	18	_		40	65	72	54	6 9	-	
G ?	Th	11	10	9	10	12	12	26	26	25	28	28	28	

Discussion

Our investigations have mainly dealt with those lactic acid bacteria whose growth was affected by calcium ions. Very little is known about the effect of these ions on the growth of lactic acid bacteria ⁹, and therefore the aim of the experiments was to determine the calcium requirements of the strains studied, and to clarify the extent to which other compounds of the basal media affect these requirements and the uptake of calcium. It was found that calcium ions could not be replaced by any of the other metal ions investigated (Mn⁺⁺, Mg⁺⁺, Fe⁺⁺, Ba⁺⁺, Sr⁺⁺). The experiments also threw light upon the effects of different concentrations of metal ions on the growth of various strains. Only magnesium ions seemed to be essential to most of the strains.

The inhibition of growth by citrate can be counteracted or at least weakened by calcium. It is of interest that the required concentrations of calcium are very low compared to the citrate concentrations. On the basis of this finding and the fact that calcium also counteracts the inhibition of growth by oxalacetate one can conclude that the counteracting effect of calcium ions is not limited to citrate alone. Moreover, it is evident that in all the basal media used calcium ions have a growth promoting effect on the strains.

The fact that the growth of about 30 of the 124 strains of lactic acid bacteria investigated were affected by calcium ions must be considered a finding of great interest. As yet, however, no conclusions can be drawn about the importance of calcium ions in the metabolism of lactic acid bacteria. The results described above indicate that calcium may play a fundamental part in the metabolism of certain organic acids.

Among the 124 investigated strains of lactobacilli there were a few which require unidentified growth factors. As yet nothing can be said about the chemical nature of these growth promoting factors although they have been separated from yeast extract and tomato juice.

The role of calcium in the metabolism of the lactic acid bacteria will be investigated further and the established unidentified growth factors will be studied to determine their chemical nature.

Summary

The nutritional requirements of 124 strains of lactic acid bacteria isolated as pure cultures from various dairy products have been studied. Some of these strains are used in cheesemaking in Finland. By growing the strains first in non-synthetic and then in synthetic media it was found that many

of them possessed very complex growth factor requirements. Several of the strains required unidentified growth factors. Particularly the Streptococcus thermophilus strains require calcium for growth. It is of interest that calcium ions counteract the inhibition of growth by citrate and some other organic anions. In order to make possible a further investigation of the effect of calcium, the efforts were largely concentrated on a search for new nutrient solutions with the simplest possible compositions. Such basal media, the chemical compositions of which must, of course, be known exactly, must first be found before a detailed examination of the mechanism of action of calcium can be undertaken.

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